

Perspective

The World Goes Bats: Living Longer and Tolerating Viruses

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For centuries, people believed that bats possessed sinister powers. Bats are thought to be ancestral hosts to many deadly viruses affecting humans including Ebola, rabies, and most recently SARS-CoV-2 coronavirus. However, bats themselves tolerate these viruses without ill effects. The second power that bats have is their longevity. Bats live much longer than similar-sized land mammals. Here we review how bats' ability to control inflammation may be contributing to their longevity. The underlying mechanisms may hold clues to developing new treatments for age-related diseases. Now may be the time to use science to exploit the secret powers of bats for human benefit.

Bat Natural History

There are more than 1,400 species of bats (Figure 1A) that inhabit all continents except Antarctica. A majority of bats are active at night and inhabit "spooky" places such as caves, wells, attics, and hollow trees, which has caused people to (falsely) attribute devilish properties to bats. Many bats live in large colonies, which promotes transmission of viruses and other pathogens. Bats are diverse in their feeding habits. Many bats feed on flying insects, others feed on fruits and nectar, but there are also carnivorous bats that catch fish and small crustaceans such as scorpions and, of course, vampire bats that feed on blood. Bats are the only species of mammal capable of powered flight, and some species can reach speeds of up to 100 miles per hour, making them the fastest mammals on earth (McCracken et al., 2016).

Bats play important role in many ecosystems. Insectivorous bats provide pest control by consuming large quantities of insects. Bats suppress crop pests and decrease pesticide use in farming (Maine and Boyles, 2015). Bats also consume mosquitoes that spread human diseases. Furthermore, many plants depend on fruit bats for pollination and seed dispersal.

One of the most amazing properties of bats is their longevity. Many bat species such as little brown bat, Brandt's bat, mouse-eared bat, and Indian flying fox have maximum lifespans of 30–40 years (Tacutu et al., 2018). Other bat species have maximum lifespans around 20 years, which is still very long for species of this size (Tacutu et al., 2018). In general, species maximum lifespan correlates positively with body mass (Austad and Fischer, 1991). Larger species tend to be longer lived. However, all bats fall above the regression line for mammals as they live longer than other mammals of similar size (Austad and Fischer, 1991; Healy et al., 2014) (Figure 1B).

Resistance to Viruses

As mentioned earlier, bats live in large dense colonies and they stay around for many years, which creates an ideal ground for

transmitting pathogens. Indeed, bats are believed to be ancestral hosts for many deadly viruses including rabies, Ebola, Marburg, Nipah, and Hendra viruses (reviewed in Banerjee et al., 2020a). The recent zoonotic transmissions of SARS-CoV (Li et al., 2005), MERS-CoV (Anthony et al., 2017), and SARS-CoV-2 (Lu et al., 2020; Zhou et al., 2020) coronaviruses are believed to have originated in bats, or were transmitted by bats to other species and then passed on to humans. Remarkably, these viruses are tolerated by bats and do not cause clinical symptoms. Even experimental inoculation of bats with some of the deadliest viruses only produced subclinical infections (Halpin et al., 2011; Middleton et al., 2007; Munster et al., 2016; Paweska et al., 2016; Schuh et al., 2017). What biological mechanisms make bats so special that they can tolerate such scourges and innocently transmit them to humans (Figure 2)?

It is important to point out that bats are a very diverse group, so a mechanism found in one species does not necessarily apply to all bats. Here, we will discuss these adaptations as they apply to individual species and the common trends that emerge. The innate immune response is the first line of defense against viruses. Cells express pattern recognition receptors, such as Toll-like receptors, that recognize pathogen-associated molecular patterns originating from viruses, bacteria, or parasites. These receptors then initiate signaling cascades that lead to expression of antiviral and proinflammatory cytokines. Antiviral cytokines include interferons (IFN), which activate expression of downstream genes that inhibit viral replication or induce death of infected cells.

Bats have a robust interferon response to RNA viruses. In the Australian black flying fox *Pteropus alecto*, IFN regulatory factor 7 (IRF7) is expressed in a wider range of tissues compared to other mammals (Zhou et al., 2014). In multiple bat species, IFN regulatory factor 3 (IRF3) has evolved a potential novel phosphorylation site, S185, that enhances activation of the downstream IFN response (Banerjee et al., 2020b). Bats also constitutively express IFN- α (Zhou et al., 2016), which in other



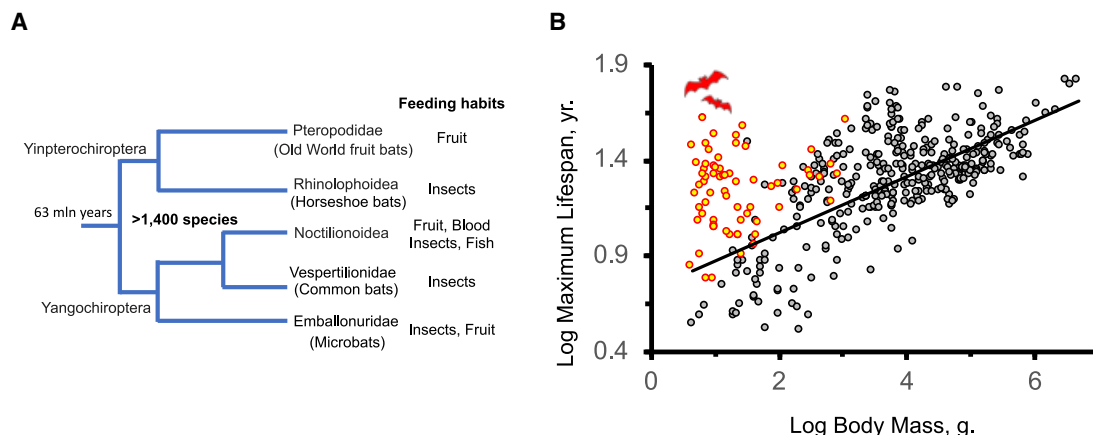


Figure 1. Bats Live Longer Than Similar-Sized Land Mammals

(A) Major lineages of bats.

(B) Relationship between lifespan and body mass in mammals. Bats are indicated by red circles; all other species of mammals are indicated by black circles. The lifespan and body mass data are from Healy et al. (2014).

mammals would lead to widespread inflammation. However, bats have evolved unique adaptations that counteract inflammation. For instance, they show significantly dampened activation of the NLRP3 inflammasome in primary immune cells compared to their human or mouse counterparts (Ahn et al., 2019). NLRP3 is an important sensor that recognizes both endogenous cellular damage and infections of either bacterial or viral origin. NLRP3 over-activation has been linked to the inflammatory state and to age-related diseases (Guo et al., 2015; Youm et al., 2013). Remarkably, dampened NLRP3 activity was observed in distant bat species, such as the fruit bat *P. alecto* and an insectivorous bat *Myotis davidii* (Ahn et al., 2019), suggesting that this is a common mechanism across species. Treating kidney cells from the big brown bat *Eptesicus fuscus* and human with poly(I:C), a viral double-stranded RNA surrogate, showed that although cells from both species induced expression of IFN- β , only human cells expressed tumor necrosis factor α (TNF- α) (Banerjee et al., 2017), a cell signaling protein involved in systemic inflammation. Analysis of the TNF promoter in the big brown bat revealed a potential repressor c-REL binding motif (Banerjee et al., 2017). Thus, downregulated TNF- α expression may be yet another strategy bats use to suppress inflammation.

Whole-genome sequencing revealed that several genes involved in innate immunity, including c-REL and NLRP3, are under positive selection in *P. alecto* and *Myotis davidii* (Zhang et al., 2013). Bats may also express unique sets of interferon stimulated genes (ISGs). Some of these genes, such as Myxovirus resistance 1 (Mx1), evolved under positive selection in bats and are reported to reduce viral replication when expressed in human cells (Fuchs et al., 2017).

Although bats mount a strong response to RNA viruses, they exhibit remarkably dampened DNA sensing. The entire PYHIN gene family was found to be missing in 10 bat species, including both fruit- and insect-eating bats (Ahn et al., 2016; Zhang et al., 2013). This gene family includes cytoplasmic DNA sensors AIM2 and IFI16 that activate the inflammasome and interferon pathways (reviewed in Schattgen and Fitzgerald, 2011). AIM2 recognizes bacterial and host DNA in the cytoplasm (Muruve et al., 2008),

forming the AIM2 inflammasome that mediates maturation of proinflammatory cytokines (IL-1 β and IL-18) (reviewed in Broz and Dixit, 2016). Although loss of the PYHIN locus appears to be universal, genomic analysis of the remaining sequences suggests different evolutionary processes leading to gene loss, rather than a single ancestral event (Ahn et al., 2016). Additionally, bats have dampened STING-dependent IFN activation (Xie et al., 2018). Upon binding to cytosolic DNA, cGAS binds and activates STING, leading to its phosphorylation on S358. Phosphorylated STING ultimately triggers the type I IFN response. Remarkably, an analysis of 30 bat species revealed that while the S358 residue is absolutely conserved among all known non-bat mammalian STING proteins, none of the bat STING proteins retain S358, leading to a weakened IFN response.

The TLR9 receptor, which preferentially recognizes DNA, appears to have evolved under purifying selection in bats and contains multiple mutations in the ligand-binding domain (Escalera-Zamudio et al., 2015). TLR9 in bats shows reduced activation by CpG-containing oligonucleotides compared to human TLR9 (Banerjee et al., 2017). Interestingly, the DNA repair protein DNA-PK, which also serves as cytoplasmic DNA sensor, is positively selected in bats (Zhang et al., 2013), possibly compensating for the lack of other DNA sensors.

Macrophages from the greater mouse-eared bat, *Myotis myotis*, challenged with various inflammatory stimuli upregulated interferon β (INF- β), TNF, and interleukin-1 β (IL-1 β). However, in bat macrophages, but not mice, this antiviral, proinflammatory response was associated with high-level transcription of the anti-inflammatory cytokine IL-10, which may help neutralize proinflammatory responses (Kacprzyk et al., 2017).

The picture that emerges from the large number of studies of bat immunity is somewhat counterintuitive. A majority of immune system adaptations found in bats dampen the immune response rather than activating it. We can learn from bats that to co-exist with viruses, controlling inflammation is more important than ramping up the immune system to combat the virus, which could lead to an inflammatory cytokine storm, exacerbating disease phenotypes and contributing to mortality. Perhaps in certain

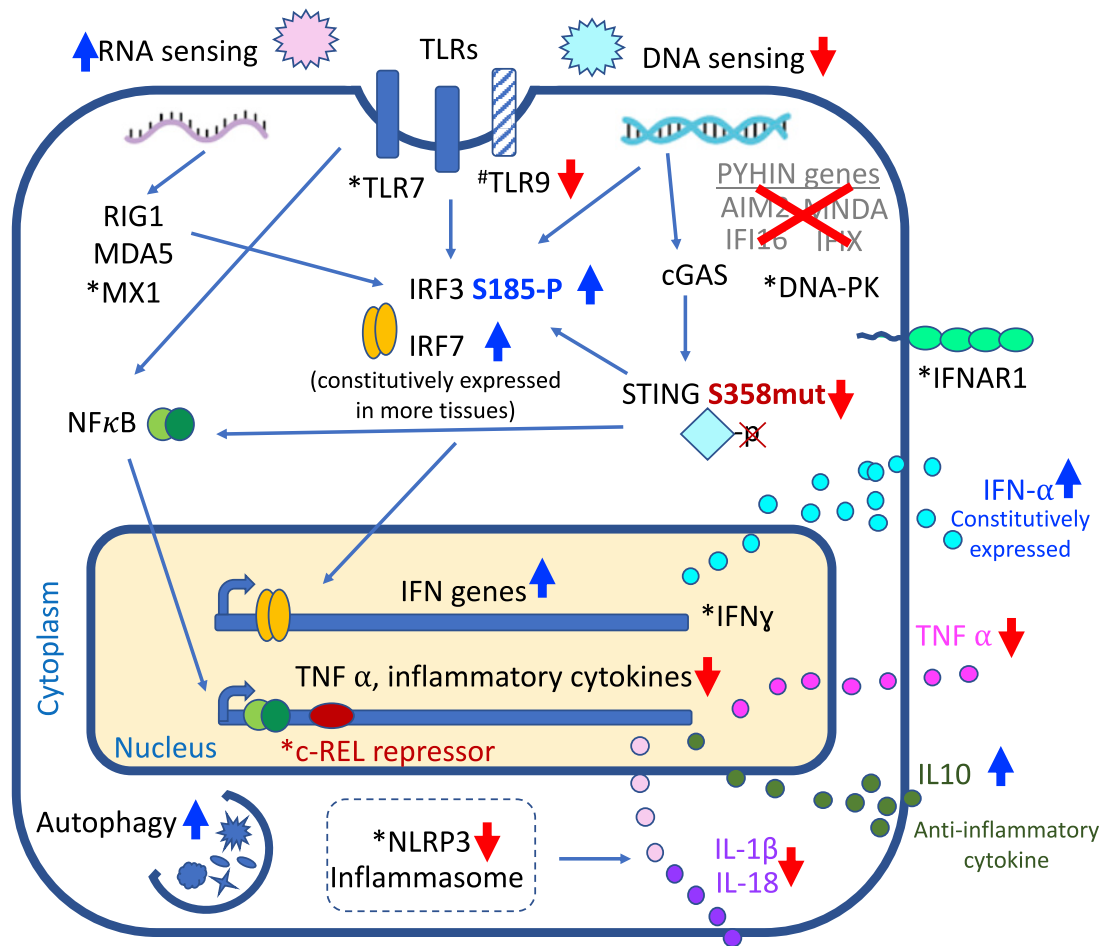


Figure 2. Unique Aspects of Immunity in Bats

The innate immune response senses cytoplasmic DNA and foreign RNA, responding by upregulation of interferon genes and production of TNF α and other inflammatory cytokines. Bat species have undergone evolutionary adaptations whereby sensing of cytoplasmic DNA is dampened through loss of PYHIN genes and a regulatory site mutation in STING. This leads to reduced expression of TNF α and other inflammatory cytokines, while anti-inflammatory cytokines like IL-10 are upregulated. Additionally, activation of the NLRP3 inflammasome is dampened in bats, leading to reduced production of IL-1 β and IL-18. Several aspects of RNA sensing are enhanced in bats. Bats also constitutively express IFN- α , show enhanced expression of interferon genes, and have more active autophagy. Taken together, these adaptations allow bats to tolerate viruses while keeping inflammatory response in check. Processes enhanced in bats are marked by upward blue arrows, while processes dampened in bats are marked by downward red arrows. *Genes under positive selection in bats. #Genes under purifying selection in bats.

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Outside of the immune system, autophagy is a process that may help bats fight viruses. Autophagy, or “self-eating,” is a cellular process that destroys and recycles cellular proteins and organelles (Saha et al., 2018). Autophagy can remove damaged proteins and can also help rid the cell of viruses. Cells from the black flying fox *P. alecto* are less susceptible than human cells to death induced by Australian bat lyssavirus, a virus related to rabies. *P. alecto* cells showed elevated basal autophagic levels, which are further induced in response to high doses of virus (Laing et al., 2019).

Why did bats evolve such tolerance to viruses? It has been proposed that unique evolution of immunity in bats is driven by flight (Banerjee et al., 2020a; Kacprzyk et al., 2017; O’Shea

et al., 2014; Zhang et al., 2013). Bats are the only flying mammals and flight requires metabolic adaptation to sudden surges in activity, rapid increases in body temperature (O’Shea et al., 2014), and perhaps, dealing with the molecular damage, such as misfolded proteins and damaged DNA, that arise (Banerjee et al., 2020a). Thus, bats may have downregulated their inflammatory pathways in order not to suffer from bouts of inflammation every time they fly. Flight is indeed a unique feature of bats, and it is plausible that some of the adaptations are related to flight. However, the fastest evolving genes, in general, are genes related to the host-pathogen arms race (Lazzaro and Clark, 2012). This evolution is driven by the presence of pathogens and happens much faster than evolution of such complex functions as flight, which require major changes to body structures. We speculate that the driving force in the evolution of bat immunity has been their lifestyle, which promotes rapid transmission of viruses. Many bat species live in gigantic colonies, where individuals

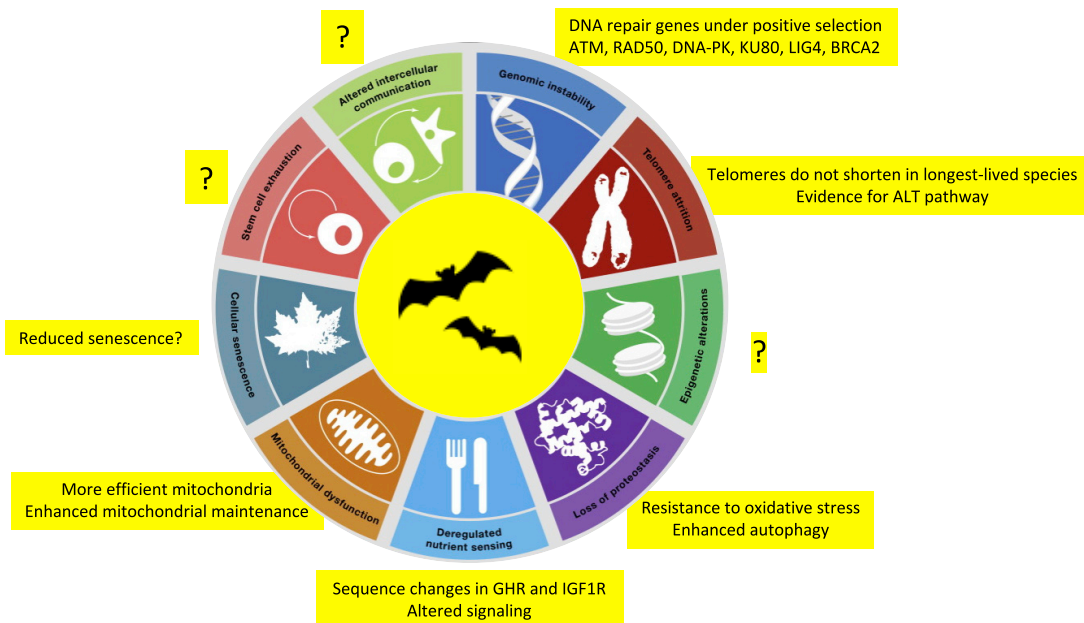


Figure 3. Longevity Adaptations in Bats

Many hallmarks of aging are altered in bats (yellow rectangles), any of which could underlie their enhanced longevity. Importantly, other hallmarks such as intercellular communication, stem cell exhaustion, and epigenetic alterations are yet to be interrogated. Modified from López-Otín et al. (2013).

spend resting periods hanging very close together on a cave ceiling or in a tree. Bat colony size may range from a few individuals to hundreds of thousands. This is the highest density among mammals, with the exception perhaps of humans in large metropolises, and considering their high mobility and foraging behavior, bats are exposed to an exceptionally high variety of viruses.

Longevity Adaptations

The long lifespan of bats is quite striking. A recent study by Wilkinson and Adams was able to reconstruct longevity across a wide range of bat species, proposing that the ancestral bat species likely lived 2.6 times longer than a similar-sized placental mammal (Wilkinson and Adams, 2019). Moreover, extreme longevity has arisen at least four separate times during bat speciation, with the longest-lived bat on record, the Brandt's bat, surviving 41 years (Podlutzky et al., 2005). These findings suggest that long lifespan is no accident; it either arose because long lifespan has fitness benefits for bats or because some other phenotype is selected that also precipitates longevity, one of which being a dampened immune response.

The reasons behind the long lifespan of bats remain debated, with scientists developing hypotheses based either on evolutionary life history or molecular studies testing known longevity pathways. Bats have several features that would favor selection for low mortality rates, including small litters (Barclay and Harder, 2003; Racey and Entwistle, 1999; Speakman, 2008), the capacity of flight (which permits escape from predators; Austad and Fischer, 1991; Holmes and Austad, 1994), and (in many species) the ability to hibernate, or enter into a low-energy torpor state. Torpor is linked to longevity in bats and other species (Turbill et al., 2011; Turbill and Prior, 2016; Wilkinson and South, 2002) and may protect the animal from bouts of starvation and/or pro-

mote homeostatic maintenance during periods of low metabolic rate. Consistent with a beneficial role for hibernation, other species that can enter hibernation, such as gray mouse lemurs and 13-lined squirrels, have longer lifespan than mice of similar size (Al-Attar and Storey, 2020). In the Wilkinson and Adams study, enhanced longevity was associated with body mass, hibernation, and cave use, which presumably provides a safer environment (Wilkinson and Adams, 2019).

Recent reviews have described pathways modulating aging, termed hallmarks or pillars (Figure 3) (Kennedy et al., 2014; López-Otín et al., 2013). The overlapping pathways these reviews describe are all intimately linked to aging in a variety of eukaryotic models. Yet it remains unclear to what extent each contributes to different aspects of aging and whether they are connected in a hierarchical structure. Almost certainly they are connected in some kind of homeostatic network that functions to maintain health in the face of damage that accrues with age. One piece of evidence for this is that genetic, dietary, or pharmacological interventions that delay aging typically affect many or all aging pillars. While an intervention likely has a direct target, the net effect is network preservation, and this affects many or all pillars. If this concept extends across species, then long-lived bats may expect to have alterations in many longevity pillars. Early evidence suggests that this is the case, and here we describe evidence connecting longevity to specific hallmarks of aging.

Genomic studies have pointed to some longevity clues. For instance, the genome of the Brandt's bat and several other species has a mutation in the growth hormone receptor gene that may interfere with transmembrane domain function (Seim et al., 2013). Growth hormone receptor hypomorphic mutations are associated with protection from diabetes and cancer in humans and long lifespan in mice (Coschigano et al., 2000; David

et al., 2011; Guevara-Aguirre et al., 2011). Indeed, bats have physiologic (e.g., pancreatic structure) and transcriptomic changes that resemble growth hormone receptor knockout mice (Liu et al., 2004; Seim et al., 2013; Swindell, 2007). There are also intriguing changes in the transmembrane region of the IGF1 receptor, which is associated with longevity in a range of model organisms and in centenarians (Kim and Lee, 2019; Seim et al., 2013). Both of these hormonal signaling pathways are intimately linked to nutrient signaling, one of the most robust pillars of aging (Figure 3). A more extensive study compared genomic data from 26 bat species to that of naked mole rats and a variety of other mammals (Davies et al., 2014). While other genomic changes were identified in both the growth hormone receptor and the IGF receptor, no evidence for positive selection was found in either gene, causing the authors to speculate that enhanced longevity was in large part linked to other genetic differences.

Genome maintenance is an important longevity assurance mechanism and another recognized pillar of aging (reviewed in Gorbunova et al., 2007). In an 8-year longitudinal study of blood samples from free-living greater mouse-eared bats (*Myotis myotis*), Huang et al. reported that DNA repair and DNA damage signaling pathways are maintained throughout lifespan, consistent with the low levels of cancer in bat species (Huang et al., 2016, 2019). Among DNA repair pathways, DNA double-strand break repair shows the strongest correlation with longevity (Tian et al., 2019). Remarkably several DNA double-strand break repair genes such as ATM, Rad50, Lig4, DNA-PK, Ku80, and BRCA2 were shown to be under positive selection in two species of bats (Zhang et al., 2013). Interestingly, some of these DNA repair genes, such as DNA-PK and Rad50, also function as DNA sensors in innate immune response (Burleigh et al., 2020; Ferguson et al., 2012; Kondo et al., 2013; Roth et al., 2014). Hence, the genetic changes that evolved in bats may modulate both processes simultaneously and the innate immune response may be an evolutionary driver of positive selection. However, further studies are needed to test whether positive selection on DNA-PK and Rad50 increases or decreases their activity as immune sensors. These changes may either compensate for dampened STING signaling or, alternatively, further decrease DNA sensing in bats.

Mitochondrial dysfunction is a feature of aging across the evolutionary spectrum and another highly supported pillar of aging (Figure 3) (Pulliam et al., 2013). Energetic demands associated with flight in bats require enhanced mitochondrial respiratory metabolism, which is expected to generate excess oxidative damage. To counteract this damage, bats have evolved more efficient mitochondria, producing less H₂O₂ per unit oxygen consumed (Brunet-Rossini, 2004). Bat fibroblasts have also been shown to have lower levels of oxidative damage to proteins and to be resistant to acute oxidative stress (Salmon et al., 2009). Limited data are available on antioxidants *in vivo* (Dammann, 2017), with superoxide dismutase (SOD) levels reported to be similar in the brain but with a trend toward higher levels in the heart. A different study compared two bats with different lifespans, finding that the longer-lived species (*Desmodus rotundus*) had higher levels of antioxidant activity across multiple tissues than its shorter-lived counterpart (*Myotis velifer*) (Conde-Pérezprina et al., 2012). This corresponded to reduced levels of DNA damage.

To help maintain proteostasis upon oxidative stress, bats express major heat shock proteins at higher levels (Chionh et al., 2019). This may simultaneously permit bats to endure high temperatures with flight and maintain protein homeostasis with age. Bats also exhibit enhanced autophagy activity with advancing age (Huang et al., 2019), suggesting that their cells are better able to clear damaged proteins and organelles. This latter finding is consistent with studies in bat fibroblasts, which detect higher levels of macroautophagy (Pride et al., 2015). Interestingly, enhanced autophagy also provides bats with enhanced protection against viruses, as described above (Laing et al., 2019).

Increased mitochondrial oxidative stress would also be expected to generate mitochondrial DNA alterations, or heteroplasmy. However, oxidative lesions in *M. myotis* are found only at low rates in an age-independent manner, suggesting better repair or removal of damaged mitochondria (Jebb et al., 2018). One recent study suggests that a system involved in limiting mitochondrial membrane potential by tethering of ATP-consuming kinases to mitochondrial membranes may be a mechanism that limits reactive oxygen species production. This system is lost in mice with age but is maintained in Seba's short-tailed bat (*Carollia perspicillata*) (Vyssokikh et al., 2020).

As they age, bats avoid upregulation of genes involved in chronic inflammation, which is typically not observed in mammals (Huang et al., 2019). This likely results from the multitude of mechanisms that evolved to suppress inflammation due to viral infections discussed above. Microbiome studies indicate that *Myotis myotis* may have stable microbiome composition that does not change over time in contrast to mice and humans, where the microbiome undergoes significant changes with age (Hughes et al., 2018). As aging-related gut dysbiosis triggers inflammation (reviewed in Kim and Jazwinski, 2018), the ability of bats to maintain stable microbiome may contribute to the lack of age-related inflammation, or by contrast, low levels of inflammation may promote a more stable microbiome. One major unanswered question is the extent to which cell senescence occurs with age in bats. Since cell senescence may be a major driver of chronic inflammation during mammalian aging, it will be important to determine whether cell senescence, another pillar, is altered with age in bat species. In-depth studies are needed to address this question *in vivo* and in cell culture.

Among other hallmarks of aging, telomere attrition has been addressed to a limited extent, with mixed results. The shorter-lived bat species, *Rhinolophus ferrumequinum* and *Miniopterus schreibersii*, do exhibit telomere shortening, but no evidence was found in the longest-lived species, *Myotis myotis* (Foley et al., 2018). This bat apparently does not express telomerase but exhibits differential expression of genes involved in telomere maintenance and the alternative lengthening of telomeres (ALT) pathway (Foley et al., 2018). Other pillars of aging, including epigenetic alterations, intercellular communication, and adult stem cell exhaustion, remain unaddressed and are also fertile grounds for further research.

Inflammation and Longevity

A majority of the mechanisms that have evolved to protect bats from viruses likely contribute to their longevity (Figure 4). Bats evolved multiple strategies to combat inflammation (discussed above), such as dampened NLRP3 inflammasome activity (Ahn

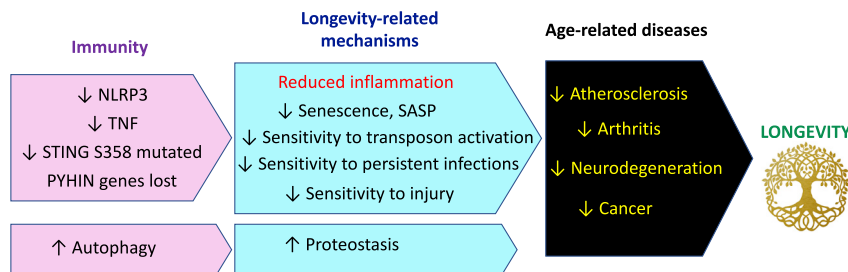


Figure 4. Changes in Immune Function in Bats that Can Promote Longevity

While many hallmarks of aging are altered in bats, we speculate that altered immunity, which leads to virus tolerance and reduced inflammation, underlies the longevity changes. The downstream consequences of the changes likely include a reduction in age-related diseases, which are fueled by inflammation such as atherosclerosis, arthritis, neurodegeneration, and cancer. This, in turn, promotes longevity in bats.

et al., 2019), dampened TNF signaling (Banerjee et al., 2017), and a dampened response to cytoplasmic DNA (Ahn et al., 2016; Xie et al., 2018; Zhang et al., 2013). Inflammation has emerged as a driver of multiple age-related pathologies, including cardiovascular diseases, cancer, Alzheimer's disease, and diabetes. This led to the concept of inflammaging, defined as the long-term result of the chronic physiological stimulation of the innate immune system, which becomes damaging during aging (Franceschi et al., 2018). Factors that trigger inflammaging include viruses, microbiome bacteria, senescent cells, and self-products of cellular damage such as debris containing cellular DNA and proteins. Reducing inflammation due to any of these factors can be beneficial for longevity; however, bat evolution seems to have attenuated mechanisms of cytoplasmic DNA sensing specifically.

Free DNA is not supposed to be in the cytoplasm, and when it does go there it signals alarm. Cytoplasmic DNA sensors have evolved to protect the cell from invading viruses. However, viruses are not the only source of cytoplasmic DNA. Mitochondrial DNA can be recognized by pattern recognition receptors such as TLR9, absent in melanoma 2 (AIM2) and cGAS and trigger activation of interferon response (reviewed in Riley and Tait, 2020; West and Shadel, 2017). Mitochondrial DNA-associated inflammasome activation has been linked to such age-related conditions as atherosclerosis (Tumurkhuu et al., 2016) and macular degeneration (Dib et al., 2015). Mitochondrial DNA release into cytoplasm has been linked to neurodegeneration and Parkinson's disease. Interestingly, in Parkin- or Pink-deficient mice mitochondrial DNA release was precipitated by exercise, while blocking INF- α receptors or deleting STING rescued inflammation (Sliter et al., 2018). Other types of mitochondrial stress, such as oxidative damage and endoplasmic reticulum stress, may also result in release of mitochondrial DNA. Thus, bats displaying improved mitochondrial maintenance and dampened cytoplasmic DNA sensing may be protected from these age-related conditions.

Recent studies demonstrated that nuclear DNA can find its way into the cytoplasm during aging and cellular senescence. In senescent cells, fragments of chromosomal DNA are expelled into the cytoplasm (Dou et al., 2017). Furthermore, nuclear DNA contains endogenous parasites in the form of transposable elements, some of which originated from viruses in the evolutionary past. Long interspersed nuclear elements (LINE1s) are retrotransposable elements that comprise 17% of the human genome. LINE1s encode a reverse-transcriptase enzyme and transpose via a copy-and-paste mechanism involving an RNA intermediate. Recent studies demonstrated that LINE1s become de-repressed in aged tissues of humans and mice, leading to

increased transcription, followed by reverse transcription and formation of cytoplasmic DNA copies of LINE1 (De Cecco et al., 2019; Simon et al., 2019). These cytoplasmic DNAs trigger the IFN response via activation of the cGAS/STING pathway, providing a new pathway for age-related sterile inflammation (De Cecco et al., 2019; Simon et al., 2019). The cGAS/STING pathway has also been implicated in cellular senescence, further linking senescence to cytoplasmic DNA (Glück et al., 2017; Yang et al., 2017a). In addition to aging, activation of retrotransposable elements and reactivity to DNA have also been associated with a variety of autoimmune conditions in humans (reviewed in Volkman and Stetson, 2014).

Remarkably, bats are unique in their ability to tolerate DNA transposable elements (Pritham and Feschotte, 2007). DNA transposons move in the genome via a cut-and-paste mechanism involving DNA intermediates. Such transposons are found in invertebrates but are generally inactivated and fossilized in the genomes of mammals. Only the vespertilionid family of bats is known to harbor significant levels of active DNA transposable elements. This bat family includes genus *Myotis*, which contains the longest-lived bats and is discussed in several contexts above, which suggests that these animals are exceptionally healthy. The ability to tolerate active DNA transposons is likely linked to dampened cytoplasmic DNA sensing.

Increasing evidence links cGAS/STING sensor to aging and age-related disease. In mouse models, STING deficiency alleviated phenotypes of prion-induced neurodegeneration and neuroinflammation (Nazmi et al., 2019). Premature aging syndrome ataxia-telangiectasia (A-T) caused by mutation in ATM kinase involved in DNA damage signaling is characterized by cancer predisposition, radiosensitivity, and neurodegeneration. Both A-T patients and *Atm*^{-/-} animal models have higher serum levels of proinflammatory cytokines and other inflammatory markers than control subjects (Westbrook and Schiestl, 2010). Neuroinflammation in A-T is associated with activation of STING and AIM2, and STING inhibition blocks the overproduction of neurotoxic cytokines (Song et al., 2019). Interestingly, it has also been reported that progerin expression in Hutchinson-Gilford syndrome fibroblasts, which is known to confer genome instability and replication stress, leads to cytoplasmic DNA and STING activation (Kreienkamp et al., 2018).

Strikingly, in a recent study of an aging Polish cohort, a SNP leading to reduced STING expression was associated with protection from age-related disease, particularly for chronic lung disease (Hamann et al., 2019), where inflammation is a major factor. The same SNP is also reported to be protective from obesity-induced cardiovascular disease (Hamann et al., 2020).



Figure 5. Adaptation to High Population Density and Air Travel

Humans evolved to live in small family groups. The first cities were settled 3000 to 4000 B.C., but significant numbers of people moved to live in large cities only over the last several centuries. Air travel is an even more recent development and became available in the last 100 years. Bats, however, have evolved to deal with these challenges over the course of 60–70 million years.

In summary, while STING activation is linked to age-related conditions, dampened STING signaling seems to confer protection from age-related pathologies in both bats and humans.

The PYHIN gene family is completely lost in multiple bat lineages (Ahn et al., 2016; Zhang et al., 2013). While PYHINs are reported to be upregulated in astrocytes and glial cells during neuroinflammation (Cox et al., 2015), the role of the PYHIN gene family in aging and age-related pathologies needs to be explored in more detail.

NLRP3 inflammasome activity is dampened in bats (Ahn et al., 2019). NLRP3 activates caspase-1 to mediate processing proinflammatory cytokines. The NLRP3 inflammasome has been linked to metabolic and cognitive diseases, including obesity, type 2 diabetes mellitus, and Alzheimer's disease (Choi and Rytter, 2014). Mice lacking NLRP3 are resistant to developing Parkinson's disease (Yan et al., 2015). IL-18, a product of inflammasome activation, may have crucial roles in the initiation and progression of atherosclerosis (Düewell et al., 2010). Multiple studies have found that NLRP3- and/or caspase-1-deficient mice show improved glucose tolerance and insulin sensitivity when exposed to a high-fat diet (Lee et al., 2013; Stienstra et al., 2011; Vandanmagsar et al., 2011; Wen et al., 2011), implicating NLRP3 in type 2 diabetes. Remarkably, ablation of the NLRP3 inflammasome in mice extends the healthspan and attenuates multiple age-related degenerative changes in the brain and in peripheral tissues (Youm et al., 2013). These studies suggest the NLRP3 inflammasome as an upstream target that controls age-related inflammation and that lowering NLRP3 activity may delay multiple age-related diseases driven by inflammation, contributing to the long lifespan of bats and serving as a target for lifespan and healthspan extension in humans.

Bats show dampened TNF- α expression in response to inflammatory stimuli (Banerjee et al., 2017). TNF- α is implicated in a multitude of disease conditions, with enhanced activity linked to a range of age-related diseases. TNF- α expression plays a direct role in obesity-linked insulin resistance (Hotamisligil et al., 1993). Activation of TNF- α is reported to contribute to disease progression in atherosclerosis (Zhang et al., 2009);

neurodegenerative conditions such as Alzheimer's disease, Huntington's disease, Parkinson's disease, and amyotrophic lateral sclerosis (Frankola et al., 2011; Greig et al., 2004; Yang et al., 2017b); osteoarthritis (Malemud et al., 2003); sarcopenia (Fan et al., 2016); and a range of other conditions. Thus, reduced TNF- α response may contribute to bat longevity.

In the continuing arms race against pathogens, evolutionary fitness requires a functional immune system. However, a highly active immune system may increase fitness in young age but limit longevity. One example is the ApoE4 allele in humans that improves pathogen resistance in children but is associated with significantly increased risk of dementia and atherosclerosis in the elderly (reviewed in Abondio et al., 2019). Why did bat evolution result in adjusted immune system functions in a way that favors longevity? We speculate that bats' exceptionally high exposure to viral pathogens forced them to develop ways to co-exist with viruses rather than to fight them. Bats are unique among mammals in the size and density of their colonies, and in their ability to fly long distances, a trait that further increases pathogen exposure (Hayman et al., 2013). Modern humans living in large metropolises and enjoying air travel may be coming close to the bat level of viral exposure. However, humans have only been enjoying this lifestyle for less than 100 years, while bats evolved 60–70 million years ago (Figure 5). We speculate that human immune systems have not evolved to deal with such frequent exposure to diverse viruses, which may manifest in a rise in autoimmune diseases and our susceptibility to pandemics. It would take an unacceptable amount of time for humans to naturally evolve adaptations similar to those of bats that would allow us to live long and healthy in these conditions; however, pharmacological interventions might be developed to bring bat-like adaptations to humans in a shorter time frame, reducing disease burden due to viral infections and delaying age-associated diseases.

Treatments Based on Bat Strategies

Bats have evolved multiple mechanisms to suppress inflammation, in particular by dampening nucleic acid sensing pathways. A number of pharmacological interventions targeting nucleic

Table 1. Bat Adaptations that Reduce Inflammation and Human Treatments

Bat Adaptations	Human Treatments	Indications	
		Current	Future
Dampened Inflammation	Target, Drug		
Dampened TNF	TNF, TNFR antagonists	autoimmune diseases	<i>age-related diseases</i>
Dampened NLRP3	NLRP3 inflammasome inhibitors		autoimmune diseases and <i>age-related diseases</i>
Dampened STING signaling	STING inhibitors and cGAS inhibitors		autoimmune diseases and <i>age-related diseases</i>
	chloroquine	malaria	
	aspirin	inflammation, fever, and <i>age-related diseases</i>	
Deleted PYHIN genes	inhibitors of AIM2, IFI16, IFIX, and MNDA		autoimmune diseases and <i>age-related diseases</i>
Tolerance of transposons	HIV reverse transcriptase, nucleoside analogs, LINE1 reverse transcriptase, nuclease, ORF1 protein	HIV, other viral infections	<i>age-related diseases</i>
Reduced senescence?	senolytics	<i>age-related diseases</i>	

acid sensing pathways had already been developed. Historically, the focus has been on developing activators of these pathways to serve as antiviral or anticancer drugs (reviewed in [Vanpouille-Box et al., 2019](#)). However, with the realization that inflammation contributes to a wide range of diseases from autoimmunity to age-related conditions, the interest has shifted to developing antagonists of nucleic acid sensors ([Sheridan, 2019](#)). This proved to be a challenging task due to high level of redundancy within nucleic acid sensing pathways and the danger of increasing vulnerability to infections. Here, the information obtained from the studies of bats can assist in drug discovery ([Table 1](#)).

TNF signaling is inhibited in bats ([Banerjee et al., 2017](#)) and anti-TNF therapies are successfully used in the treatment of a wide variety of autoimmune diseases such as rheumatoid arthritis, psoriasis, ulcerative colitis, and Crohn disease (reviewed in [Croft and Siegel, 2017](#)). Anti-TNF therapeutics are typically monoclonal antibodies to either TNF or TNF receptors. These drugs effectively alleviate inflammation, prevent tissue destruction, and reduce patient mortality ([Monaco et al., 2015](#)). Interestingly, arthritis patients treated with TNF antagonists also showed improvements in cardiovascular conditions ([Tam et al., 2014](#)), suggesting that TNF inhibition may confer a broader benefit for age-related diseases.

NLRP3 inflammasome is dampened in bats ([Ahn et al., 2019](#)), and aberrant activation of NLRP3 is linked to a wide range of age-related diseases. Several direct NLRP3 inhibitors have been reported that alleviate inflammatory conditions in mice ([Coll et al., 2015](#); [He et al., 2014](#); [Jiang et al., 2017](#)). Many new inhibitors with improved potency and safety profiles are being developed to treat inflammatory conditions including age-related diseases such as Alzheimer's, Parkinson's, and cardiovascular diseases (reviewed in [Zahid et al., 2019](#)).

Bats show a dampened cGAS-STING pathway ([Xie et al., 2018](#)). Remarkably, recent progress in drug development has been achieved precisely in targeting these two proteins (reviewed in [Ding et al., 2020](#)). Small molecule inhibitors of STING have recently been reported ([Haag et al., 2018](#); [Li et al., 2018](#)). These molecules reduced STING-mediated inflammatory cytokine pro-

duction in both human and mouse cells and were effective in ameliorating inflammatory conditions in mouse models ([Haag et al., 2018](#)). cGAS inhibitors are also being developed ([Lama et al., 2019](#)). The first application of these compounds will be for the treatment of autoimmune conditions, but ultimately regimens may be developed where these compounds target aging and age-related diseases. Remarkably, several old and widely used drugs have been found to attenuate cGAS. This includes aspirin ([Dai et al., 2019](#)) and antimalarial drugs quinacrine and chloroquine ([An et al., 2015](#); [Piscianz et al., 2018](#)), supporting the idea that targeting pathways inhibited in bats confers health benefits to humans. Interestingly, aspirin is widely used, although controversially, in small doses to reduce cardiovascular disease risk ([Ittaman et al., 2014](#); [McNeil et al., 2018](#)), and it has been shown to extend lifespan modestly in male mice ([Strong et al., 2008](#)).

Bats may also help identify novel therapeutic targets. As discussed above, bats lost the entire PYHIN family of genes responsible for nucleic acid sensing ([Ahn et al., 2016](#); [Zhang et al., 2013](#)). It will be interesting to explore pharmacological targeting of PYHIN genes, such as AIM2, IFI16, and others, for the treatment of age-related and autoimmune conditions. Interestingly, AIM2-deficient mice do not show severe adverse phenotypes, but display enhanced resistance to ionizing radiation ([Hu et al., 2016](#)).

Recent studies showed that retrotransposons become activated with age and drive sterile inflammation via cGAS-STING pathway ([De Cecco et al., 2019](#); [Simon et al., 2019](#)). Bats are also particularly good at handling transposable elements ([Pritham and Feschotte, 2007](#)). Remarkably, inhibiting retrotransposition activity using nucleoside reverse-transcriptase inhibitors developed to target HIV reverse transcriptase ameliorated age-related inflammation in mouse models ([De Cecco et al., 2019](#); [Simon et al., 2019](#)). Mice showed reduced expression of inflammatory cytokines; lower expression of the p16^{NK4A} gene, which is a marker of cellular senescence and aging; and reduced methylation age. Treatment of progeroid SIRT6-deficient mice with nucleoside reverse-transcriptase inhibitors doubled the mouse lifespan, reduced inflammation, and alleviated pathology across multiple tissues ([Simon et al., 2019](#)). Therefore, targeting retrotransposable elements may be a promising strategy to ameliorate

age-related inflammation. In addition to existing nucleoside analogs that serve as chain terminators for generic reverse transcriptases, specific inhibitors targeting LINE1 reverse transcriptase or endonuclease protein can be developed, which would have lower toxicity for other cellular polymerases. Such compounds may even be used to promote healthy aging.

Senescent cells have been shown to contribute to development of a variety of age-related conditions by promoting sterile inflammation (reviewed in Freund et al., 2010). Little is known about cellular senescence in bats. However, since senescence requires cGAS (Glück et al., 2017; Yang et al., 2017a), and bats dampen cGAS-STING signaling (Xie et al., 2018), one can speculate that the senescence response is attenuated in bats. Thus, senolytic compounds that selectively eliminate senescent cells (Wissler Gerdes et al., 2020) may be another way to pharmacologically promote bat-like adaptations in humans. It is already reported that genetic strategies to ablate senescent cells lead to healthspan and lifespan extension in mice (Baker et al., 2016; Jeon et al., 2017). Many different senolytic molecules have been identified (Fuhrmann-Stroissnigg et al., 2018; Kirkland and Tchkonja, 2017), and several are reported to delay aging and/or protect against age-related diseases (Chang et al., 2016; Xu et al., 2018; Yousefzadeh et al., 2018).

Future Perspectives

In summary, besides serving as a source of deadly diseases and harboring viruses similar to SARS-CoV-2, which caused the current pandemics, bats have a lot to offer humanity by illuminating the pathways to develop novel therapeutics to treat age-related conditions and promote longevity. Already studies of the altered innate immune responses in bats point to several classes of small molecules, some of which have links to aging. However, we can only see the tip of the iceberg when it comes to understanding how bats deal with inflammation, and clearly more studies are warranted. This is also true for other hallmarks of aging, which are still minimally explored. In addition to finding small molecules targeting specific pathways that can be tested in humans, it will be of interest to engineer specific bat alterations in mice and determine whether this leads to enhanced lifespan and healthspan.

Bats have evolved skewed, and ultimately successful, strategies to experience longer and healthier lives, even if this is a secondary outcome of selection for responses to viral infections and/or the dramatic range of metabolic states that accompany periods of flight and torpor. Humans in the last century have created a lifestyle that has gone bats; we live in high densities and (many of us) travel extensively, enhancing exposure to and spread of pathogens. By embracing “batty” strategies to deal with the challenges that our new lifestyle presents, we may be able to solve what look to be the two biggest medical challenges of the 21st century: the rise of viral pandemics and the ever-increasing prevalence of chronic diseases that all share aging as their biggest risk factor.

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REFERENCES

- Abondio, P., Sazzini, M., Garagnani, P., Boattini, A., Monti, D., Franceschi, C., Luiselli, D., and Giuliani, C. (2019). The genetic variability of *APOE* in different human populations and its implications for longevity. *Genes (Basel)* 10, 222.
- Ahn, M., Cui, J., Irving, A.T., and Wang, L.F. (2016). Unique loss of the PYHIN gene family in bats amongst mammals: implications for inflammasome sensing. *Sci. Rep.* 6, 21722.
- Ahn, M., Anderson, D.E., Zhang, Q., Tan, C.W., Lim, B.L., Luko, K., Wen, M., Chia, W.N., Mani, S., Wang, L.C., et al. (2019). Dampened NLRP3-mediated inflammation in bats and implications for a special viral reservoir host. *Nat. Microbiol.* 4, 789–799.
- Al-Attar, R., and Storey, K.B. (2020). Suspended in time: molecular responses to hibernation also promote longevity. *Exp. Gerontol.* 134, 110889.
- An, J., Woodward, J.J., Sasaki, T., Minie, M., and Elkon, K.B. (2015). Cutting edge: antimalarial drugs inhibit IFN- β production through blockade of cyclic GMP-AMP synthase-DNA interaction. *J. Immunol.* 194, 4089–4093.
- Anthony, S.J., Gilardi, K., Menachery, V.D., Goldstein, T., Ssebide, B., Mbabazi, R., Navarrete-Macias, I., Liang, E., Wells, H., Hicks, A., et al. (2017). Further evidence for bats as the evolutionary source of Middle East respiratory syndrome coronavirus. *MBio* 8, e00373-17.
- Austad, S.N., and Fischer, K.E. (1991). Mammalian aging, metabolism, and ecology: evidence from the bats and marsupials. *J. Gerontol.* 46, B47–B53.
- Baker, D.J., Childs, B.G., Durik, M., Wijers, M.E., Sieben, C.J., Zhong, J., Saltness, R.A., Jeganathan, K.B., Verzosa, G.C., Pezeshki, A., et al. (2016). Naturally occurring p16(Ink4a)-positive cells shorten healthy lifespan. *Nature* 530, 184–189.
- Banerjee, A., Rapin, N., Bollinger, T., and Misra, V. (2017). Lack of inflammatory gene expression in bats: a unique role for a transcription repressor. *Sci. Rep.* 7, 2232.
- Banerjee, A., Baker, M.L., Kulcsar, K., Misra, V., Plowright, R., and Mossman, K. (2020a). Novel insights into immune systems of bats. *Front. Immunol.* 11, 26.
- Banerjee, A., Zhang, X., Yip, A., Schulz, K.S., Irving, A.T., Bowdish, D., Golding, B., Wang, L.F., and Mossman, K. (2020b). Positive selection of a serine residue in bat IRF3 confers enhanced antiviral protection. *iScience* 23, 100958.
- Barclay, R.M.R., and Harder, L.D. (2003). Life histories of bats: life in the slow lane. In *Ecology of Bats*, T.H. Kunz and M.B. Fenton, eds. (Chicago University Press), pp. 209–253.
- Broz, P., and Dixit, V.M. (2016). Inflammasomes: mechanism of assembly, regulation and signalling. *Nat. Rev. Immunol.* 16, 407–420.
- Brunet-Rossini, A.K. (2004). Reduced free-radical production and extreme longevity in the little brown bat (*Myotis lucifugus*) versus two non-flying mammals. *Mech. Ageing Dev.* 125, 11–20.
- Burleigh, K., Maltbaek, J.H., Cambier, S., Green, R., Gale, M., Jr., James, R.C., and Stetson, D.B. (2020). Human DNA-PK activates a STING-independent DNA sensing pathway. *Sci. Immunol.* 5, eaba4219.
- Chang, J., Wang, Y., Shao, L., Laberge, R.M., Demaria, M., Campisi, J., Janakiraman, K., Sharpless, N.E., Ding, S., Feng, W., et al. (2016). Clearance of senescent cells by ABT263 rejuvenates aged hematopoietic stem cells in mice. *Nat. Med.* 22, 78–83.
- Chionh, Y.T., Cui, J., Koh, J., Mendenhall, I.H., Ng, J.H.J., Low, D., Itahana, K., Irving, A.T., and Wang, L.F. (2019). High basal heat-shock protein expression in bats confers resistance to cellular heat/oxidative stress. *Cell Stress Chaperones* 24, 835–849.
- Choi, A.J., and Ryter, S.W. (2014). Inflammasomes: molecular regulation and implications for metabolic and cognitive diseases. *Mol. Cells* 37, 441–448.
- Coll, R.C., Robertson, A.A., Chae, J.J., Higgins, S.C., Muñoz-Planillo, R., In-serra, M.C., Vetter, I., Dungan, L.S., Monks, B.G., Stutz, A., et al. (2015). A small-molecule inhibitor of the NLRP3 inflammasome for the treatment of inflammatory diseases. *Nat. Med.* 21, 248–255.
- Conde-Pérezprina, J.C., Luna-López, A., González-Puertos, V.Y., Zenteno-Savín, T., León-Galván, M.A., and Königsmberg, M. (2012). DNA MMR systems, microsatellite instability and antioxidant activity variations in two species of

wild bats: *Myotis velifer* and *Desmodus rotundus*, as possible factors associated with longevity. *Age (Dordr.)* 34, 1473–1492.

Coschigano, K.T., Clemmons, D., Bellush, L.L., and Kopchick, J.J. (2000). Assessment of growth parameters and life span of GHR/BP gene-disrupted mice. *Endocrinology* 147, 2608–2613.

Cox, D.J., Field, R.H., Williams, D.G., Baran, M., Bowie, A.G., Cunningham, C., and Dunne, A. (2015). DNA sensors are expressed in astrocytes and microglia in vitro and are upregulated during gliosis in neurodegenerative disease. *Glia* 63, 812–825.

Croft, M., and Siegel, R.M. (2017). Beyond TNF: TNF superfamily cytokines as targets for the treatment of rheumatic diseases. *Nat. Rev. Rheumatol.* 13, 217–233.

Dai, J., Huang, Y.J., He, X., Zhao, M., Wang, X., Liu, Z.S., Xue, W., Cai, H., Zhan, X.Y., Huang, S.Y., et al. (2019). Acetylation blocks cGAS activity and inhibits self-DNA-induced autoimmunity. *Cell* 176, 1447–1460.e14.

Dammann, P. (2017). Slow aging in mammals—lessons from African mole-rats and bats. *Semin. Cell Dev. Biol.* 70, 154–163.

David, A., Hwa, V., Metherell, L.A., Netchine, I., Camacho-Hübner, C., Clark, A.J., Rosenfeld, R.G., and Savage, M.O. (2011). Evidence for a continuum of genetic, phenotypic, and biochemical abnormalities in children with growth hormone insensitivity. *Endocr. Rev.* 32, 472–497.

Davies, K.T., Tsagkogeorga, G., Bennett, N.C., Dávalos, L.M., Faulkes, C.G., and Rossiter, S.J. (2014). Molecular evolution of growth hormone and insulin-like growth factor 1 receptors in long-lived, small-bodied mammals. *Gene* 549, 228–236.

De Cecco, M., Ito, T., Petrashen, A.P., Elias, A.E., Skvir, N.J., Criscione, S.W., Caligiana, A., Brocculi, G., Adney, E.M., Boeke, J.D., et al. (2019). L1 drives IFN in senescent cells and promotes age-associated inflammation. *Nature* 566, 73–78.

Dib, B., Lin, H., Maidana, D.E., Tian, B., Miller, J.B., Bouzika, P., Miller, J.W., and Vavvas, D.G. (2015). Mitochondrial DNA has a pro-inflammatory role in AMD. *Biochim. Biophys. Acta* 1853 (11 Pt A), 2897–2906.

Ding, C., Song, Z., Shen, A., Chen, T., and Zhang, A. (2020). Small molecules targeting the innate immune cGAS–STING–TBK1 signaling pathway. *Acta Pharm. Sin. B*. Published online March 13, 2020. <https://doi.org/10.1016/j.apsb.2020.03.001>.

Dou, Z., Ghosh, K., Vizioli, M.G., Zhu, J., Sen, P., Wangenstein, K.J., Smithy, J., Lan, Y., Lin, Y., Zhou, Z., et al. (2017). Cytoplasmic chromatin triggers inflammation in senescence and cancer. *Nature* 550, 402–406.

Duewell, P., Kono, H., Rayner, K.J., Sirois, C.M., Vladimer, G., Bauernfeind, F.G., Abela, G.S., Franchi, L., Núñez, G., Schnurr, M., et al. (2010). NLRP3 inflammasomes are required for atherogenesis and activated by cholesterol crystals. *Nature* 464, 1357–1361.

Escalera-Zamudio, M., Zepeda-Mendoza, M.L., Loza-Rubio, E., Rojas-Anaya, E., Méndez-Ojeda, M.L., Arias, C.F., and Greenwood, A.D. (2015). The evolution of bat nucleic acid-sensing Toll-like receptors. *Mol. Ecol.* 24, 5899–5909.

Fan, J., Kou, X., Yang, Y., and Chen, N. (2016). MicroRNA-regulated proinflammatory cytokines in sarcopenia. *Mediators Inflamm.* 2016, 1438686.

Ferguson, B.J., Mansur, D.S., Peters, N.E., Ren, H., and Smith, G.L. (2012). DNA-PK is a DNA sensor for IRF-3-dependent innate immunity. *eLife* 1, e00047.

Foley, N.M., Hughes, G.M., Huang, Z., Clarke, M., Jebb, D., Whelan, C.V., Petit, E.J., Touzalin, F., Farcy, O., Jones, G., et al. (2018). Growing old, yet staying young: the role of telomeres in bats' exceptional longevity. *Sci. Adv.* 4, e0926.

Franceschi, C., Garagnani, P., Parini, P., Giuliani, C., and Santoro, A. (2018). Inflammaging: a new immune-metabolic viewpoint for age-related diseases. *Nat. Rev. Endocrinol.* 14, 576–590.

Frankola, K.A., Greig, N.H., Luo, W., and Tweedie, D. (2011). Targeting TNF- α to elucidate and ameliorate neuroinflammation in neurodegenerative diseases. *CNS Neurol. Disord. Drug Targets* 10, 391–403.

Freund, A., Orjalo, A.V., Desprez, P.Y., and Campisi, J. (2010). Inflammatory networks during cellular senescence: causes and consequences. *Trends Mol. Med.* 16, 238–246.

Fuchs, J., Hölzer, M., Schilling, M., Patzina, C., Schoen, A., Hoenen, T., Zimmer, G., Marz, M., Weber, F., Müller, M.A., and Kochs, G. (2017). Evolution and antiviral specificities of interferon-induced Mx proteins of bats against Ebola, influenza, and other RNA viruses. *J. Virol.* 91, e00361-17.

Fuhrmann-Stroissnigg, H., Niedernhofer, L.J., and Robbins, P.D. (2018). Hsp90 inhibitors as senolytic drugs to extend healthy aging. *Cell Cycle* 17, 1048–1055.

Glück, S., Guey, B., Gulen, M.F., Wolter, K., Kang, T.W., Schmacke, N.A., Bridgeman, A., Rehwinkel, J., Zender, L., and Ablasser, A. (2017). Innate immune sensing of cytosolic chromatin fragments through cGAS promotes senescence. *Nat. Cell Biol.* 19, 1061–1070.

Gorbunova, V., Seluanov, A., Mao, Z., and Hine, C. (2007). Changes in DNA repair during aging. *Nucleic Acids Res.* 35, 7466–7474.

Greig, N.H., Mattson, M.P., Perry, T., Chan, S.L., Giordano, T., Sambamurti, K., Rogers, J.T., Ovadia, H., and Lahiri, D.K. (2004). New therapeutic strategies and drug candidates for neurodegenerative diseases: p53 and TNF-alpha inhibitors, and GLP-1 receptor agonists. *Ann. N Y Acad. Sci.* 1035, 290–315.

Guevara-Aguirre, J., Balasubramanian, P., Guevara-Aguirre, M., Wei, M., Madia, F., Cheng, C.W., Hwang, D., Martin-Montalvo, A., Saavedra, J., Ingles, S., et al. (2011). Growth hormone receptor deficiency is associated with a major reduction in pro-aging signaling, cancer, and diabetes in humans. *Sci. Transl. Med.* 3, 70ra13.

Guo, H., Callaway, J.B., and Ting, J.P. (2015). Inflammasomes: mechanism of action, role in disease, and therapeutics. *Nat. Med.* 21, 677–687.

Haag, S.M., Gulen, M.F., Reymond, L., Gibelin, A., Abrami, L., Decout, A., Heymann, M., van der Goot, F.G., Turcatti, G., Behrendt, R., and Ablasser, A. (2018). Targeting STING with covalent small-molecule inhibitors. *Nature* 559, 269–273.

Halpin, K., Hyatt, A.D., Fogarty, R., Middleton, D., Bingham, J., Epstein, J.H., Rahman, S.A., Hughes, T., Smith, C., Field, H.E., and Daszak, P.; Henipavirus Ecology Research Group (2011). Pteropid bats are confirmed as the reservoir hosts of henipaviruses: a comprehensive experimental study of virus transmission. *Am. J. Trop. Med. Hyg.* 85, 946–951.

Hamann, L., Ruiz-Moreno, J.S., Szwed, M., Mossakowska, M., Lundvall, L., Schumann, R.R., Opitz, B., and Puzianowska-Kuznicka, M. (2019). STING SNP R293Q is associated with a decreased risk of aging-related diseases. *Gerontology* 65, 145–154.

Hamann, L., Szwed, M., Mossakowska, M., Chudek, J., and Puzianowska-Kuznicka, M. (2020). First evidence for STING SNP R293Q being protective regarding obesity-associated cardiovascular disease in age-advanced subjects - a cohort study. *Immun. Ageing* 17, 7.

Hayman, D.T., Bowen, R.A., Cryan, P.M., McCracken, G.F., O'Shea, T.J., Peel, A.J., Gilbert, A., Webb, C.T., and Wood, J.L. (2013). Ecology of zoonotic infectious diseases in bats: current knowledge and future directions. *Zoonoses Public Health* 60, 2–21.

He, Y., Varadarajan, S., Muñoz-Planillo, R., Burberry, A., Nakamura, Y., and Núñez, G. (2014). 3,4-methylenedioxy- β -nitrostyrene inhibits NLRP3 inflammasome activation by blocking assembly of the inflammasome. *J. Biol. Chem.* 289, 1142–1150.

Healy, K., Guillerme, T., Finlay, S., Kane, A., Kelly, S.B., McClean, D., Kelly, D.J., Donohue, I., Jackson, A.L., and Cooper, N. (2014). Ecology and mode-of-life explain lifespan variation in birds and mammals. *Proc. Biol. Sci.* 281, 20140298.

Holmes, D.J., and Austad, S.N. (1994). Fly now, die later: life-history correlates of gliding and flying in mammals. *J. Mammal.* 75, 224–226.

Hotamisligil, G.S., Shargill, N.S., and Spiegelman, B.M. (1993). Adipose expression of tumor necrosis factor-alpha: direct role in obesity-linked insulin resistance. *Science* 259, 87–91.

Hu, B., Jin, C., Li, H.B., Tong, J., Ouyang, X., Cetinbas, N.M., Zhu, S., Strowig, T., Lam, F.C., Zhao, C., et al. (2016). The DNA-sensing AIM2 inflammasome controls radiation-induced cell death and tissue injury. *Science* 354, 765–768.

Huang, Z., Jebb, D., and Teeling, E.C. (2016). Blood miRNomes and transcriptomes reveal novel longevity mechanisms in the long-lived bat, *Myotis myotis*. *BMC Genomics* 17, 906.

Huang, Z., Whelan, C.V., Foley, N.M., Jebb, D., Touzalin, F., Petit, E.J., Puechmaile, S.J., and Teeling, E.C. (2019). Longitudinal comparative transcriptomics reveals unique mechanisms underlying extended healthspan in bats. *Nat. Ecol. Evol.* **3**, 1110–1120.

Hughes, G.M., Leech, J., Puechmaile, S.J., Lopez, J.V., and Teeling, E.C. (2018). Is there a link between aging and microbiome diversity in exceptional mammalian longevity? *PeerJ* **6**, e4174.

Ittaman, S.V., VanWormer, J.J., and Rezkalla, S.H. (2014). The role of aspirin in the prevention of cardiovascular disease. *Clin. Med. Res.* **12**, 147–154.

Jebb, D., Foley, N.M., Whelan, C.V., Touzalin, F., Puechmaile, S.J., and Teeling, E.C. (2018). Population level mitogenomics of long-lived bats reveals dynamic heteroplasmy and challenges the free radical theory of ageing. *Sci. Rep.* **8**, 13634.

Jeon, O.H., Kim, C., Laberge, R.M., Demaria, M., Rathod, S., Vasserot, A.P., Chung, J.W., Kim, D.H., Poon, Y., David, N., et al. (2017). Local clearance of senescent cells attenuates the development of post-traumatic osteoarthritis and creates a pro-regenerative environment. *Nat. Med.* **23**, 775–781.

Jiang, H., He, H., Chen, Y., Huang, W., Cheng, J., Ye, J., Wang, A., Tao, J., Wang, C., Liu, Q., et al. (2017). Identification of a selective and direct NLRP3 inhibitor to treat inflammatory disorders. *J. Exp. Med.* **214**, 3219–3238.

Kacprzyk, J., Hughes, G.M., Palsson-McDermott, E.M., Quinn, S.R., Puechmaile, S.J., O'Neill, L.A.J., and Teeling, E.C. (2017). A potent anti-inflammatory response in bat macrophages may be linked to extended longevity and viral tolerance. *Acta Chiropt.* **19**, 219–228.

Kennedy, B.K., Berger, S.L., Brunet, A., Campisi, J., Cuervo, A.M., Epel, E.S., Franceschi, C., Lithgow, G.J., Morimoto, R.I., Pessin, J.E., et al. (2014). Geroscience: linking aging to chronic disease. *Cell* **159**, 709–713.

Kim, S., and Jazwinski, M.S. (2018). The gut microbiota and healthy aging: a mini-review. *Gerontology* **64**, 513–520.

Kim, S.S., and Lee, C.K. (2019). Growth signaling and longevity in mouse models. *BMB Rep.* **52**, 70–85.

Kirkland, J.L., and Tchkonja, T. (2017). Cellular senescence: a translational perspective. *EBioMedicine* **27**, 21–28.

Kondo, T., Kobayashi, J., Saitoh, T., Maruyama, K., Ishii, K.J., Barber, G.N., Komatsu, K., Akira, S., and Kawai, T. (2013). DNA damage sensor MRE11 recognizes cytosolic double-stranded DNA and induces type I interferon by regulating STING trafficking. *Proc. Natl. Acad. Sci. USA* **110**, 2969–2974.

Kreienkamp, R., Graziano, S., Coll-Bonfill, N., Bedia-Diaz, G., Cybulla, E., Vindigni, A., Dorsett, D., Kubben, N., Batista, L.F.Z., and Gonzalo, S. (2018). A cell-intrinsic interferon-like response links replication stress to cellular aging caused by progerin. *Cell Rep.* **22**, 2006–2015.

Laing, E.D., Sterling, S.L., Weir, D.L., Beauregard, C.R., Smith, I.L., Larsen, S.E., Wang, L.F., Snow, A.L., Schaefer, B.C., and Broder, C.C. (2019). Enhanced autophagy contributes to reduced viral infection in black flying fox cells. *Viruses* **11**, 260.

Lama, L., Adura, C., Xie, W., Tomita, D., Kamei, T., Kuryavii, V., Gogakos, T., Steinberg, J.I., Miller, M., Ramos-Espiritu, L., et al. (2019). Development of human cGAS-specific small-molecule inhibitors for repression of dsDNA-triggered interferon expression. *Nat. Commun.* **10**, 2261.

Lazzaro, B.P., and Clark, A. (2012). Rapid evolution of innate immune response genes. In *Rapidly Evolving Genes and Genetic Systems*, R.S. Singh, J. Xu, and R.J. Kulathinal, eds. (Oxford Scholarship Online).

Lee, H.M., Kim, J.J., Kim, H.J., Shong, M., Ku, B.J., and Jo, E.K. (2013). Upregulated NLRP3 inflammasome activation in patients with type 2 diabetes. *Diabetes* **62**, 194–204.

Li, W., Shi, Z., Yu, M., Ren, W., Smith, C., Epstein, J.H., Wang, H., Cramer, G., Hu, Z., Zhang, H., et al. (2005). Bats are natural reservoirs of SARS-like coronaviruses. *Science* **310**, 676–679.

Li, S., Hong, Z., Wang, Z., Li, F., Mei, J., Huang, L., Lou, X., Zhao, S., Song, L., Chen, W., et al. (2018). The cyclopeptide astin C specifically inhibits the innate immune CDN sensor STING. *Cell Rep.* **25**, 3405–3421.e7.

Liu, J.L., Coschigano, K.T., Robertson, K., Lipssett, M., Guo, Y., Kopchick, J.J., Kumar, U., and Liu, Y.L. (2004). Disruption of growth hormone receptor gene

causes diminished pancreatic islet size and increased insulin sensitivity in mice. *Am. J. Physiol. Endocrinol. Metab.* **287**, E405–E413.

López-Otín, C., Blasco, M.A., Partridge, L., Serrano, M., and Kroemer, G. (2013). The hallmarks of aging. *Cell* **153**, 1194–1217.

Lu, R., Zhao, X., Li, J., Niu, P., Yang, B., Wu, H., Wang, W., Song, H., Huang, B., Zhu, N., et al. (2020). Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding. *Lancet* **395**, 565–574.

Maine, J.J., and Boyles, J.G. (2015). Bats initiate vital agroecological interactions in corn. *Proc. Natl. Acad. Sci. USA* **112**, 12438–12443.

Malemud, C.J., Islam, N., and Haqqi, T.M. (2003). Pathophysiological mechanisms in osteoarthritis lead to novel therapeutic strategies. *Cells Tissues Organs* **174**, 34–48.

McCracken, G.F., Safi, K., Kunz, T.H., Dechmann, D.K., Swartz, S.M., and Winkelski, M. (2016). Airplane tracking documents the fastest flight speeds recorded for bats. *R. Soc. Open Sci.* **3**, 160398.

McNeil, J.J., Wolfe, R., Woods, R.L., Tonkin, A.M., Donnan, G.A., Nelson, M.R., Reid, C.M., Lockery, J.E., Kirpach, B., Storey, E., et al.; ASPREE Investigator Group (2018). Effect of aspirin on cardiovascular events and bleeding in the healthy elderly. *N. Engl. J. Med.* **379**, 1509–1518.

Middleton, D.J., Morrissy, C.J., van der Heide, B.M., Russell, G.M., Braun, M.A., Westbury, H.A., Halpin, K., and Daniels, P.W. (2007). Experimental Nipah virus infection in pteropid bats (*Pteropus poliocephalus*). *J. Comp. Pathol.* **136**, 266–272.

Monaco, C., Nanchahal, J., Taylor, P., and Feldmann, M. (2015). Anti-TNF therapy: past, present and future. *Int. Immunol.* **27**, 55–62.

Munster, V.J., Adney, D.R., van Doremalen, N., Brown, V.R., Miazgowiec, K.L., Milne-Price, S., Bushmaker, T., Rosenke, R., Scott, D., Hawkinson, A., et al. (2016). Replication and shedding of MERS-CoV in Jamaican fruit bats (*Artibeus jamaicensis*). *Sci. Rep.* **6**, 21878.

Muruve, D.A., Pétrilli, V., Zaiss, A.K., White, L.R., Clark, S.A., Ross, P.J., Parks, R.J., and Tschopp, J. (2008). The inflammasome recognizes cytosolic microbial and host DNA and triggers an innate immune response. *Nature* **452**, 103–107.

Nazmi, A., Field, R.H., Griffin, E.W., Haugh, O., Hennessy, E., Cox, D., Reis, R., Tortorelli, L., Murray, C.L., Lopez-Rodriguez, A.B., et al. (2019). Chronic neurodegeneration induces type I interferon synthesis via STING, shaping microglial phenotype and accelerating disease progression. *Glia* **67**, 1254–1276.

O’Shea, T.J., Cryan, P.M., Cunningham, A.A., Fooks, A.R., Hayman, D.T., Luis, A.D., Peel, A.J., Plowright, R.K., and Wood, J.L. (2014). Bat flight and zoonotic viruses. *Emerg. Infect. Dis.* **20**, 741–745.

Paweska, J.T., Storm, N., Grobbelaar, A.A., Markotter, W., Kemp, A., and Jansen van Vuren, P. (2016). Experimental inoculation of Egyptian fruit bats (*Rousettus aegyptiacus*) with Ebola virus. *Viruses* **8**, 29.

Piscianz, E., Cuzzoni, E., Sharma, R., Tesser, A., Sapra, P., and Tommasini, A. (2018). Reappraisal of antimalarials in interferonopathies: new perspectives for old drugs. *Curr. Med. Chem.* **25**, 2797–2810.

Podlitsky, A.J., Khritankov, A.M., Ovodov, N.D., and Austad, S.N. (2005). A new field record for bat longevity. *J. Gerontol. A Biol. Sci. Med. Sci.* **60**, 1366–1368.

Pride, H., Yu, Z., Sunchu, B., Mochnick, J., Coles, A., Zhang, Y., Buffenstein, R., Hornsby, P.J., Austad, S.N., and Pérez, V.I. (2015). Long-lived species have improved proteostasis compared to phylogenetically-related shorter-lived species. *Biochem. Biophys. Res. Commun.* **457**, 669–675.

Pritham, E.J., and Feschotte, C. (2007). Massive amplification of rolling-circle transposons in the lineage of the bat *Myotis lucifugus*. *Proc. Natl. Acad. Sci. USA* **104**, 1895–1900.

Pulliam, D.A., Bhattacharya, A., and Van Remmen, H. (2013). Mitochondrial dysfunction in aging and longevity: a causal or protective role? *Antioxid. Redox Signal.* **19**, 1373–1387.

Racey, P.A., and Entwistle, A.C. (1999). Life history and reproductive strategies of bats. In *Reproductive Biology of Bats*, P.H. Krutzsch and E.G. Crichton, eds. (Academic Press).

- Riley, J.S., and Tait, S.W. (2020). Mitochondrial DNA in inflammation and immunity. *EMBO Rep.* *21*, e49799.
- Roth, S., Rottach, A., Lotz-Havla, A.S., Laux, V., Muschwack, A., Gersting, S.W., Muntau, A.C., Hopfner, K.P., Jin, L., Vanness, K., et al. (2014). Rad50-CARD9 interactions link cytosolic DNA sensing to IL-1 β production. *Nat. Immunol.* *15*, 538–545.
- Saha, S., Panigrahi, D.P., Patil, S., and Bhutia, S.K. (2018). Autophagy in health and disease: a comprehensive review. *Biomed. Pharmacother.* *104*, 485–495.
- Salmon, A.B., Leonard, S., Masamsetti, V., Pierce, A., Podlutzky, A.J., Podlutzskaya, N., Richardson, A., Austad, S.N., and Chaudhuri, A.R. (2009). The long lifespan of two bat species is correlated with resistance to protein oxidation and enhanced protein homeostasis. *FASEB J.* *23*, 2317–2326.
- Schattgen, S.A., and Fitzgerald, K.A. (2011). The PYHIN protein family as mediators of host defenses. *Immunol. Rev.* *243*, 109–118.
- Schuh, A.J., Amman, B.R., Sealy, T.K., Spengler, J.R., Nichol, S.T., and Towner, J.S. (2017). Egyptian rousette bats maintain long-term protective immunity against Marburg virus infection despite diminished antibody levels. *Sci. Rep.* *7*, 8763.
- Seim, I., Fang, X., Xiong, Z., Lobanov, A.V., Huang, Z., Ma, S., Feng, Y., Turanov, A.A., Zhu, Y., Lenz, T.L., et al. (2013). Genome analysis reveals insights into physiology and longevity of the Brandt's bat *Myotis brandtii*. *Nat. Commun.* *4*, 2212.
- Sheridan, C. (2019). Drug developers switch gears to inhibit STING. *Nat. Biotechnol.* *37*, 199–201.
- Simon, M., Van Meter, M., Ablaeva, J., Ke, Z., Gonzalez, R.S., Taguchi, T., De Cecco, M., Leonova, K.I., Kogan, V., Helfand, S.L., et al. (2019). LINE1 derepression in aged wild-type and SIRT6-deficient mice drives inflammation. *Cell Metab.* *29*, 871–885.e5.
- Sliter, D.A., Martinez, J., Hao, L., Chen, X., Sun, N., Fischer, T.D., Burman, J.L., Li, Y., Zhang, Z., Narendra, D.P., et al. (2018). Parkin and PINK1 mitigate STING-induced inflammation. *Nature* *561*, 258–262.
- Song, X., Ma, F., and Herrup, K. (2019). Accumulation of cytoplasmic DNA due to ATM deficiency activates the microglial viral response system with neurotoxic consequences. *J. Neurosci.* *39*, 6378–6394.
- Speakman, J.R. (2008). The physiological costs of reproduction in small mammals. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* *363*, 375–398.
- Stienstra, R., van Diepen, J.A., Tack, C.J., Zaki, M.H., van de Veerdonk, F.L., Perera, D., Neale, G.A., Hooiveld, G.J., Hijmans, A., Vroegrijk, I., et al. (2011). Inflammasome is a central player in the induction of obesity and insulin resistance. *Proc. Natl. Acad. Sci. USA* *108*, 15324–15329.
- Strong, R., Miller, R.A., Astle, C.M., Floyd, R.A., Flurkey, K., Hensley, K.L., Javors, M.A., Leeuwenburgh, C., Nelson, J.F., Ongini, E., et al. (2008). Nordihydroguaiaretic acid and aspirin increase lifespan of genetically heterogeneous male mice. *Aging Cell* *7*, 641–650.
- Swindell, W.R. (2007). Gene expression profiling of long-lived dwarf mice: longevity-associated genes and relationships with diet, gender and aging. *BMC Genomics* *8*, 353.
- Tacutu, R., Thornton, D., Johnson, E., Budovsky, A., Barardo, D., Craig, T., Diana, E., Lehmann, G., Toren, D., Wang, J., et al. (2018). Human ageing genomic resources: new and updated databases. *Nucleic Acids Res.* *46* (D1), D1083–D1090.
- Tam, L.S., Kitas, G.D., and González-Gay, M.A. (2014). Can suppression of inflammation by anti-TNF prevent progression of subclinical atherosclerosis in inflammatory arthritis? *Rheumatology (Oxford)* *53*, 1108–1119.
- Tian, X., Firsanov, D., Zhang, Z., Cheng, Y., Luo, L., Tomblin, G., Tan, R., Simon, M., Henderson, S., Steffan, J., et al. (2019). SIRT6 is responsible for more efficient DNA double-strand break repair in long-lived species. *Cell* *177*, 622–638.e22.
- Tumurkhuu, G., Shimada, K., Dagvadorj, J., Crother, T.R., Zhang, W., Luthringer, D., Gottlieb, R.A., Chen, S., and Arditi, M. (2016). Ogg1-dependent DNA repair regulates NLRP3 inflammasome and prevents atherosclerosis. *Circ. Res.* *119*, e76–e90.
- Turbill, C., Bieber, C., and Ruf, T. (2011). Hibernation is associated with increased survival and the evolution of slow life histories among mammals. *Proc. Biol. Sci.* *278*, 3355–3363.
- Turbill, C., and Prior, S. (2016). Thermal climate-linked variation in annual survival rate of hibernating rodents: shorter winter dormancy and lower survival in warmer climates. *Funct. Ecol.* *30*, 1366–1372.
- Vandanmagsar, B., Youm, Y.H., Ravussin, A., Galgani, J.E., Stadler, K., Myrnat, R.L., Ravussin, E., Stephens, J.M., and Dixit, V.D. (2011). The NLRP3 inflammasome instigates obesity-induced inflammation and insulin resistance. *Nat. Med.* *17*, 179–188.
- Vanpouille-Box, C., Hoffmann, J.A., and Galluzzi, L. (2019). Pharmacological modulation of nucleic acid sensors - therapeutic potential and persisting obstacles. *Nat. Rev. Drug Discov.* *18*, 845–867.
- Volkman, H.E., and Stetson, D.B. (2014). The enemy within: endogenous retroelements and autoimmune disease. *Nat. Immunol.* *15*, 415–422.
- Vyssokikh, M.Y., Holtze, S., Averina, O.A., Lyamzaev, K.G., Panteleeva, A.A., Marey, M.V., Zinovkin, R.A., Severin, F.F., Skulachev, M.V., Fasel, N., et al. (2020). Mild depolarization of the inner mitochondrial membrane is a crucial component of an anti-aging program. *Proc. Natl. Acad. Sci. USA* *117*, 6491–6501.
- Wen, H., Gris, D., Lei, Y., Jha, S., Zhang, L., Huang, M.T., Brickey, W.J., and Ting, J.P. (2011). Fatty acid-induced NLRP3-ASC inflammasome activation interferes with insulin signaling. *Nat. Immunol.* *12*, 408–415.
- West, A.P., and Shadel, G.S. (2017). Mitochondrial DNA in innate immune responses and inflammatory pathology. *Nat. Rev. Immunol.* *17*, 363–375.
- Westbrook, A.M., and Schiestl, R.H. (2010). Atm-deficient mice exhibit increased sensitivity to dextran sulfate sodium-induced colitis characterized by elevated DNA damage and persistent immune activation. *Cancer Res.* *70*, 1875–1884.
- Wilkinson, G.S., and Adams, D.M. (2019). Recurrent evolution of extreme longevity in bats. *Biol. Lett.* *15*, 20180860.
- Wilkinson, G.S., and South, J.M. (2002). Life history, ecology and longevity in bats. *Aging Cell* *1*, 124–131.
- Wissler Gerdes, E.O., Zhu, Y., Tchkonja, T., and Kirkland, J.L. (2020). Discovery, development, and future application of senolytics: theories and predictions. *FEBS J.* *287*, 2418–2427.
- Xie, J., Li, Y., Shen, X., Goh, G., Zhu, Y., Cui, J., Wang, L.F., Shi, Z.L., and Zhou, P. (2018). Dampened STING-dependent interferon activation in bats. *Cell Host Microbe* *23*, 297–301.e4.
- Xu, M., Pirtskhalava, T., Farr, J.N., Weigand, B.M., Palmer, A.K., Weivoda, M.M., Inman, C.L., Ogrodnik, M.B., Hachfeld, C.M., Fraser, D.G., et al. (2018). Senolytics improve physical function and increase lifespan in old age. *Nat. Med.* *24*, 1246–1256.
- Yan, Y., Jiang, W., Liu, L., Wang, X., Ding, C., Tian, Z., and Zhou, R. (2015). Dopamine controls systemic inflammation through inhibition of NLRP3 inflammasome. *Cell* *160*, 62–73.
- Yang, H., Wang, H., Ren, J., Chen, Q., and Chen, Z.J. (2017a). cGAS is essential for cellular senescence. *Proc. Natl. Acad. Sci. USA* *114*, E4612–E4620.
- Yang, H.M., Yang, S., Huang, S.S., Tang, B.S., and Guo, J.F. (2017b). Microglial activation in the pathogenesis of Huntington's disease. *Front. Aging Neurosci.* *9*, 193.
- Youm, Y.H., Grant, R.W., McCabe, L.R., Albarado, D.C., Nguyen, K.Y., Ravussin, A., Pistell, P., Newman, S., Carter, R., Laque, A., et al. (2013). Canonical Nlrp3 inflammasome links systemic low-grade inflammation to functional decline in aging. *Cell Metab.* *18*, 519–532.
- Yousefzadeh, M.J., Zhu, Y., McGowan, S.J., Angelini, L., Fuhrmann-Stroissnigg, H., Xu, M., Ling, Y.Y., Melos, K.I., Pirtskhalava, T., Inman, C.L., et al. (2018). Fisetin is a senotherapeutic that extends health and lifespan. *EBioMedicine* *36*, 18–28.
- Zahid, A., Li, B., Kombe, A.J.K., Jin, T., and Tao, J. (2019). Pharmacological inhibitors of the NLRP3 inflammasome. *Front. Immunol.* *10*, 2538.

Zhang, H., Park, Y., Wu, J., Chen, X.p., Lee, S., Yang, J., Dellsperger, K.C., and Zhang, C. (2009). Role of TNF-alpha in vascular dysfunction. *Clin. Sci. (Lond.)* 116, 219–230.

Zhang, G., Cowled, C., Shi, Z., Huang, Z., Bishop-Lilly, K.A., Fang, X., Wynne, J.W., Xiong, Z., Baker, M.L., Zhao, W., et al. (2013). Comparative analysis of bat genomes provides insight into the evolution of flight and immunity. *Science* 339, 456–460.

Zhou, P., Cowled, C., Mansell, A., Monaghan, P., Green, D., Wu, L., Shi, Z., Wang, L.F., and Baker, M.L. (2014). IRF7 in the Australian black flying fox, *Pteropus alecto*: evidence for a unique expression pattern and functional conservation. *PLoS One* 9, e103875.

Zhou, P., Tachedjian, M., Wynne, J.W., Boyd, V., Cui, J., Smith, I., Cowled, C., Ng, J.H., Mok, L., Michalski, W.P., et al. (2016). Contraction of the type I IFN locus and unusual constitutive expression of IFN- α in bats. *Proc. Natl. Acad. Sci. USA* 113, 2696–2701.

Zhou, P., Yang, X.L., Wang, X.G., Hu, B., Zhang, L., Zhang, W., Si, H.R., Zhu, Y., Li, B., Huang, C.L., et al. (2020). A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature* 579, 270–273.